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Centre d'Etude Régional pour l'Amélioration
de l'Adaptation à la Sécheresse

**Electrolyte leakage and fatty acid profile as
indices of membrane integrity in groundnut
(*Arachis hypogaea* L.) subjected to water stress**

TECHNICAL REPORT

March 1 to May 27, 1999

Dr. Emmanuel Akubugwo
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Abia State, NIGERIA

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SUMMAKY

Membrane integrity of the leaves of three groundnut cultivars (Fleur11, GC-85 and 57-422) subjected to varying levels of water stress was assessed using electrolyte leakage and changes in fatty acid profile as indices. The effect of the treatments on leaf dry matter was also studied. To date, studies on the leaf membrane electrolyte leakage as well as dry matter have been completed while studies on the fatty acid is still in progress. Data obtained from the completed studies have been statistically analysed. Results indicate that the absolute integrity of the leaf membranes were perturbed by the treatments. Membrane damage was evident and increased in magnitude in the different cultivars with increase in the severity of the applied drought. Significant variety and water regime effects were observed in the electrolyte leakage. In the dry matter however, there were no variety effect, rather strong water regime effects were observed. In all the data analysed so far, there were no block effects.

INTRODUCTION

Groundnut is an important food and cash crop in many tropical and sub-tropical countries including Nigeria and Senegal. Like other crops, drought is a major limiting factor in its growth and production. Classical breeding techniques have been employed by breeders to select drought resistant cultivar to counter the adverse effect of the incessant water deficit problems prevalent in these regions. These techniques, though effective are time consuming. There is therefore, need to develop rapid and effective methods of screening of plants for drought resistance. It is generally accepted that biochemical methods offer a good alternative. It is known that when plants are subjected to water deficit, they respond through alterations in their biochemical make-up. These changes when properly characterised are veritable tools for screening of plants for drought tolerance. Such changes of interest have been observed in the membranes.

The fluidity of membranes (which is essential for it to function under variable conditions) is governed by the chemical composition of the membrane. Drought destroys cell membranes especially those of chloroplasts and mitochondria (Vieira da Silver et al., 1974; Pham Thi and Vieira da Silva, 1975). When the integrity of the membrane is compromised by dehydration (as occurs in drought), electrolytes leak out of the cell. This leakage can be assessed by measuring the conductivity of the solution in which they are kept (Sullivan, 1971). There exists a direct relationship between the ability of membranes to maintain fluidity and the nature of their lipid composition. It is therefore expedient that the lipid content and fatty acid composition of the leaves of plants be properly characterised especially under water stress conditions. This is so because of the critical role cell membrane stability plays in drought tolerance.

Protoplasmic tolerance involve mechanisms which include osmoregulation, control of enzymatic activities as well as control of re-arrangements in membrane structures (Losch, 1986; Turner and Jones, 1980). Reports (Ferrari-Iliou et al., 1984 ; Pham Thi et al., 1985 ; Quartacci and Navari-Izzo, 1992) indicate that many annual plants respond to drought stress by changing the polar lipid composition of their leaf cell membranes (Repellin et al., 1997), reported similar changes in the leaf membrane of young coconut palms subjected to water stress. These changes present in the form of reduction in the polar lipid content especially the glycolipids. Another feature of these modifications is decreases in the poly unsaturated fatty acid content. This effect is more pronounced in the monogalactosyl diacylglycerol (MGDG) fraction, than in the other polar lipid types. These modifications are products of the inter play of three events which simultaneously occur in the plant during water deficit, Viz.: 1/ Increased lipolytic activities (El-Hafid et al., 1989). 2/ Inhibition of lipid synthesis (Pham Thi et al., 1985, 1987), and 3/ Enhanced production of scavenging free radicals (Ferrari-Iliou et al., 1992).

Available reports indicate a good correlation between the extent of modification of the polar lipids composition of the leaf membranes and the abilities of the different cultivars within a species to tolerate and adapt to water deficit (Pham Thi et al., 1990). Leaf membrane polar lipid characteristics is considered a good index for the evaluation of protoplasmic tolerance to drought stress within a plant species (Hubac et al., 1989 ; Monteiro de Paula et al., 1990 ; Pham Thi et al., 1990 ; Chen and Liu, 1991) and could be used by breeders as a screening test for drought tolerance. The present investigation aims at correlating membrane integrity as assessed by electrolyte leakage with a parallel assessment using changes in the fatty acid profile in three peanut cultivars subjected to varying levels of water stress in the green house.

MATERIALS AND METHODS

Seed collection

Certified seeds of three peanut cultivars viz. -57-422, Fleur-I 1, and GC8-35 were obtained from CNRA/ISRA in Bambey Senegal. They were chosen because agronomically they are known to have good drought resistance capacity.

Growth Conditions

Sandy soil was used as the potting medium. The soil characteristics were as previously described by Annerose (1990). The soil (6.3kg) were packed in plastic pots of volume 3.51 (diam =15cm, ht. =20cm) with drainage holes at the bottom. The seeds were treated with the fungicide Granox (Captafol 10%, Benomyl 10% and Carbofuran 20%) to prevent fungal attack. After irrigation to field capacity, three seeds were sown per pot at a soil depth of 3 cm. Ten days after planting, the plants were thinned down to one plant per pot to obtain plants of uniform growth vigour. The soil was fertilised with N.P.K. (8-18-27) at the rate of 150kg/ha to remove nutrient deficiency as a limiting factor. The plants were kept in the green house at CERAAS, Thiès, Senegal, (longitude 14°42' ,latitude 16°57').

Soil Moisture Treatment

Four water regimes were chosen after a preliminary experiment. They are-

- 1 .Field capacity (full irrigation).
- 2.Low water stress (-1.25 to -1.54 MPa).
- 3.Medium water stress (-2.31 to -2.52 MPa).
- 4.Severe water stress (-3.15 MPa).

Experimental Design and Statistical Analysis

Block randomised design with three replicates and four watering regime was employed. A total of 144 pots in three experimental blocks separated by inter block distances of 70cm

were used. Within a **block**, there were twelve (12) plots. **One** plot is made up of four pots and **constitute** an experimental unit.

Measurements

All measurements were made on the third leaf counting from the stem apex **during** the **vegetative/early** flowering stages of the plant. Eleven hours **after** the choice of a particular stress **level** from the preliminary experiment was made, studies on the following parameters were performed.

Physiological Measurements

1. Leaf waterpotential (ψ , MPa)

This was determined in the green house using hydraulic pressure (Campbell JIG) in the apical part of the leaf. Measurements were taken at a fixed hour in the morning to avoid variations in water potential associated with time of the day.

2. Leaf dry matter determination

Fresh leaves from the plants were weighed and kept in the oven at 80° C for **48h**, re-weighed and the **difference** in weight noted.

3. Membrane electrolyte leakage

Membrane integrity was assessed at the various water potentials using electrolyte leakage method of Ames,(1983).

Biochemical Analysis

1. Extraction of membrane lipids

The Harvested second developed leaves were first fixed in boiling water for 2-5 minutes. Total lipids were extracted in chloroform: methanol : water (1:1:1,v/v/v) according to the method of Allen and Good, (1971). The lipid **extracts** were dried under a stream of **nitrogen** and stored in ethanol : toluene mixture(1:4 v/v) in the refrigerator till when required for analysis.

2. Separation of the lipids into various classes by Thin Layer chromatography (TLC)

This **was** performed according to the **procedure** described by Lepage, (1967)

3. Saponification and transmethylation of Lipids

Both the total lipid and the various lipid classes obtained after TLC was effected by acid **catalysed** transesterification of the saponified lipid.

4. Analysis of the methylesters of the fatty acids by GLC

The **Varian** GLC system equipped with flame ionisation detector (FID) was used with C- 17 as the **internal** standard while Helium was the carrier gas. The flow rate was 2ml/min while column temperature was maintained at 200°C.

RESULTS

The effect of water stress on the **absolute** integrity of the leaf membranes of the **studied** groundnut cultivars are shown in Table 1 a. In **each cultivar**, the effect was most pronounced in the severe stress. **GC5-35** was the most resistant while **Fleur-I 1** was the least resistant. This picture is shown clearer by the percent membrane **damage** in Table 1 b.

The nature of the interactions of the water stress on the electrolyte leakage in the different cultivars is shown in Table 1 c. There are significant ($P<0.05$) **differences between** the amount of electrolyte leakage in the control (Field **capacity**) and the values obtained in the various levels of applied stress. However, the **difference of the effects** of low and moderate stress were not significant ($P>0.05$). This Table also shows that amongst the varieties, the only significant ($P<0.05$) interaction occurred in **Fleur-I 1** relative to **GC8-35** only.

Table 1a. Absolute membrane integrity in three groundnut (*Arachis hypogaea* L.) cultivars subjected to different levels of water stress.

	<u>Percentage membrane integrity</u>		
	Fleur- 11	GC8-35	57-422
Field capacity	88.17a	87.02a	88.17a
Low stress	74.07b	83.55b	76.70b
Medium stress	51.21b	77.57b	75.04b
Severe stress	42.15c	66.78c	58.31c

In each column, means followed by the same letters are not significantly ($P < 0.05$). different according to Duncan's multiple range test.

Table 1 b. Percentage membrane damage associated with water stress levels in three groundnut (*A. hypogaea* L.) cultivars.

	<u>Percent membrane damage</u>		
	Fleur- 11	GC8-35	57-422
Low stress	12.71	3.55	8.63
Medium stress	16.78	10.48	9.78
Severe stress	38.85	22.88	30.56

Table 1c. Nature of interactions (variety effects) within various water regimes observed in the membrane integrity studies using electrolyte leakage method.

	Field capacity	Low stress	Moderate	Severe stress
f-leur- 11	88.17 a	74.07a	51.21a	42.15a
GC8-35	87.02 a	83.55b	77.57b	66.78b
57-422	88.17a	76.70ab	75.04ab	58.31ab

In each column. means followed by the same letters are not significantly ($P < 0.05$). different according to Duncan's multiple range test.

The effect of the water stress on biomass accumulation in the groundnut leaves is shown in Table 2. Biomass (dry matter) calculated on wet weight basis increased in the leaves with increased intensity of water deficit. GC8-35 had the highest dry matter. However, the dry matter content of 57-422 was slightly greater than that of Fleur-II. There are significant ($P<0.05$) differences between the dry matter content of the control (Field capacity) and those of the various levels of applied stress. However, the difference of the effects of low and moderate stress were not significant ($P>0.05$). The interactions observed among the varieties were very complex.

Table 2. Leaf dry matter changes associated with water stress levels in three peanut (*A. hypogea* L.) cultivars.

	<u>Leaf dry matter (g/100g wet weight).</u>		
	Fleur- 11	GC8-35	57-422
Field capacity	17.54	24.55	30.67
Low stress	38.71	33.14	35.09
Moderate stress	46.27	54.31	51.25
Severe stress	61.01	66.48	59.36

DISCUSSION

Our projections in this study are 1. Application of stress will cause membrane damage whose severity will increase with increased level of water deficit. 2. The membrane damage will vary amongst the different cultivars depending on their drought resistance capacities and 3. The pattern of the observed changes in 1 and 2 above will be the same both in the electrolyte leakage and in the fatty acid profile studies.

The results obtained showed that absolute integrity of the leaf membrane was compromised by the application of water stress. The intensity of this compromise was heightened by the severity of the stress.(Table 1a.). This result is in deed amplified by the percentage membrane damage (Table 1 b.). Various workers have shown that water stress results in membrane damage (Vieira da Silver et al.,1974; Pham Thi and Vieira da Silva, 1975).It is also accepted that measurement of the conductivity of the medium in which the damaged membranes are kept will give a true picture of the extent of the damage (Sullivan,1971).Our results are consistent with these earlier observations and also with our projection. Based on the result so far analysed, the water deficit tolerance capacity of the three cultivars is in the order Fleur-I 1> 57-422> GC8-35. If the result from the fatty acid profile studies (which is in

progress) corroborates this finding, our projections in this study would have been fully actualised.

PROBLEMS

The major problem we had in prosecuting this project was that of adapting the new equipment which CERAAS recently acquired to the local environment. A number of the methodologies also needed to be modified. Much success have been achieved in this regard. With the level of commitment shown by the staff on the ground, these issues will soon be effectively completely resolved.

PERSPECTIVES

Based on the facilities available in CERAAS for lipid research, I have the following perspectives-

1. That the centre be used to train Regional expert in the use of lipid techniques in drought research.
2. That special training workshop on lipid study techniques be mounted soon for the staff on the ground to enable them make effective use of the ultra modern equipment in CERAAS.

ON-GOING EXPERIMENT

During my stay at CERAAS, I also participated in an on going experiment titled "Root growth of cowpea genotypes in hydroponics, aeroponic and under different soil moisture conditions in the green house". Data is currently being generated from the study.

REFERENCES

- Allen, C. F. and Good P.;(1971). Acyl lipids in photosynthetic systems. *Methods in Enzymology*, 23 : 533-547.
- Ames, B. N. (1983). Assay of inorganic phosphate and phosphatase s. In :*Methods in Enzymology*, 8. S. P. Colowick and N. O. Kaplan. Acad. Press. New York, London. : 115-118.
- Chen, S Y. and Liu, J. (1991). The effect of water stress on membrane fluidity of leaf mitochondria of sugarcane and its relation to membrane lipid peroxidation. *Acta. Phytophysiol. Sin.* 17: 285-289.
- El-Hafid, L , Pham Thi, A. T. , Zuily-Fodil, Y. and Vieira da Silva, J. (1989). Enzymatic break down of polar lipids in cotton leaves under water stress. I. Degradation of monogalactosyldiacylglycerol. *Plant Physiol.*, 27: 495-502.
- Ferrari-Iliou, R. ,Pham Thi, A. T. and Vieira da Silva J. (1984). Effect of water stress on the lipid and fatty acid composition of cotton (*Gossypium hirsutum*) chloroplast. *Physiol. Plant.*, 62: 219-224.

- Ferrari-Iliou, R. Pham Thi, A. T. , Mazliak, P and Vieira da Silva. J(1992).Effets de la peroxydation des lipides membranaires sur la végétaux. Ann. Biol. 31: 1-23.
- Hubac, C. , Guerrier, D. Ferran, J. and Tremolières , A. (1989).Change of leaf lipid composition during water stress in two genotypes of *Lupinus albus* resistance or susceptible to drought . *Plant. Physiol. Biochem.* 54: 484-486.
- Losch. R. (1986). *Physiology I. Plant Water Relations. Prog. Bot.* (Springer-Verlag, Berlin and Heidelberg). 48: 56-79.
- Monteiro de Paula, F. Pham Thi, A. T. and Vieira da Silva, J. Demandre ,C. and Mazliak, P (1990).Effects of water stress on the molecular species composition of polar lipids from *Vigna unguiculata* leaves .*Plant. Sci.*, 66: 185193.
- Pham Thi, A. T. ,Borrel-Flood, C., Vieira da Silva, J. B.; Austin, A. M .and Mazlik, P.,(1987). Effects of drought on (1-14C-oleic acid and (1-C)-linolenic acid desaturation in cotton leaves *Physiol. Plant.* 69: 147-150.
- Pham Thi, A. T. and Vieira da Silva, J. B. (1975).Action d'un traitement osmotique sur l'ultrastructure des feuilles de cotonnier (*Gossypium hirsutum* L. and *G Anomalum* Waw. Et Peyr). *C. R. Acad. Sci., Paris*, 280 : 2857-2860.
- Pham Thi., A. T. ,Borrel-Flood. C., Vieira da Silva. J. , Justin. A. M. and Mazliak, P. (1985) Effect of water stress on lipid metabolism in cotton leaves. *Phytochemistry.* 24: 723-727.
- Pham Thi, A, T. and Vieira da Silva, and J Mazliak, P(1990).The role of membrane lipids in drought resistance of plants. *Bull. Soc. Bot. Fr.* 137: 99-144.
- Quartacci,M. F. and Nvaari-Izzo, F. (1992). Water stress and free radical mediated changes in sunflower seedlings. *J. Plant. Physiol.* **139**:621-625.
- Repellin, A. Pham Thi, A T, Tashakorie, A., Sasah, Y. Daniel ,C. and Zuily-Fodil, Y.(1997). Leaf membrane lipids and drought tolerance in young coconut palms (*Cocos nucifera* L.). *Eur J Agron.* **6**:25-33.
- Turner, N. C. and Jones, M. M. (1980). Turgor maintenance by osmotic adjustment: a review and evaluation. In: Turner, N. C. and Kramer P. J. (Eds.).Adaptation of plants to water and High temperature stress. Wiley Interscience. New York. : 87-104.
- Vieira da Silva, J.B., Naylor, A. W. and Kramer, P. J.,(1974). Some ultrastructural and enzymatic effect of water stress in cotton (*Gossypium hirsutum* L.) leaves. *Proc Nat. Acad. Sci. USA* : 3243-3247.

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